

Double Collection PCR Kit

Before you begin:

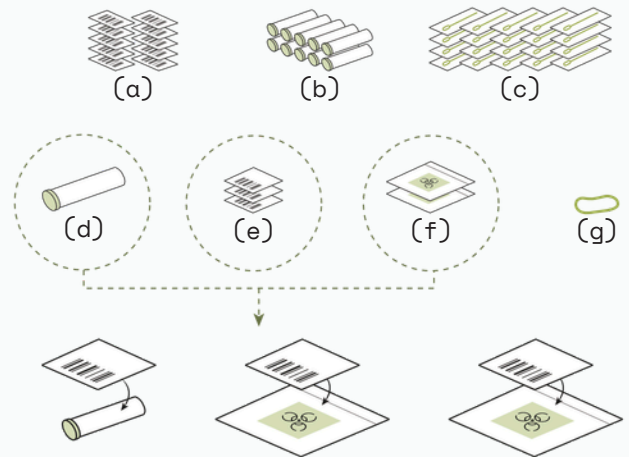
Please read through this instructions sheet before you begin sample collection. Samples that are not properly registered, collected or labeled may not be usable.

Bulk double collect sample kit contents

- (50x) biohazard bags
- (200x) 10mL tubes
- (400x) swabs
- (75x) pool barcode labels
- (25x) 50mL tubes
- (200x) individual barcode labels

01 Unbox and prepare kit contents

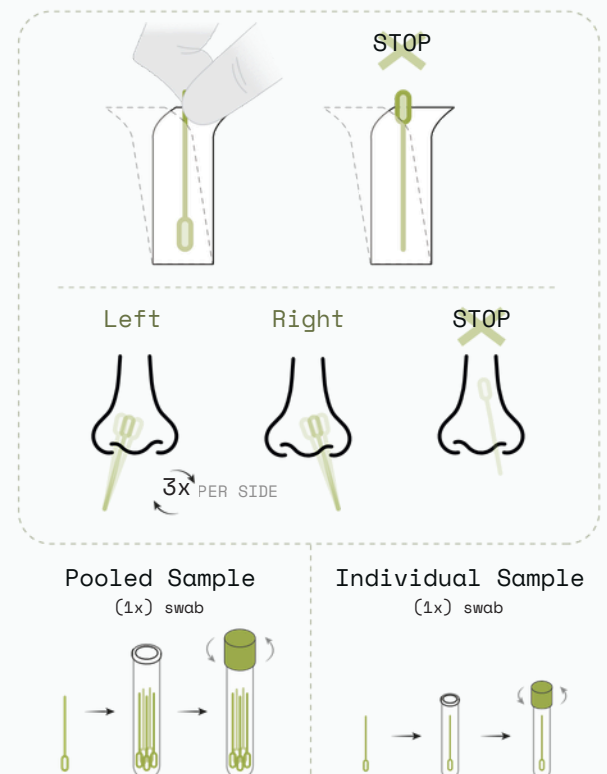
1. Wash & dry or sanitize your hands.
2. To prepare materials for **1 pool**, remove from the shipping box:
 - a. (10x) sequentially numbered “individual” barcode labels
 - b. (10x) 10mL tubes
 - c. (20x) swabs
 - d. (1x) 50mL tube
 - e. (1x) set of 3 matching “pool” barcode labels
 - f. (2x) biohazard bags
 - g. (1x) rubber band (not included in test kit)
3. Place (1x) barcode on each of the following:
 - (1x) Pooled Sample Tube (large 50mL tube)
 - (2x) biohazard bags
4. Unscrew the top of the large 50mL tube and place it upright in a stable container or tube rack.



02 Collect samples

1. As participants approach the testing station, ensure they each provide **2 samples**:
 - (1x) for pooled collection
 - (1x) for individual tests
2. Check to ensure the 3 matching barcode labels are placed on the 50mL tube, and (2x) biohazard bags.
3. Participants will collect their own sample; do not swab anyone.
4. Give each participant one (1) polyester nasal swab. Participants should remove the swab from the package without touching the polyester tip.
5. Instruct the participant to insert the tip of the swab into their nostril until the tip is no longer visible.
6. Participants should then rotate the swab in a circle around the entire inside edge of each nostril at least (3x) times and can remove the swab from their nostril upon completion.
7. Once removed, add the participant’s swab to a 10 ml tube. **10ml tubes should never have more than one swab. Tightly seal each cap.**
8. Have participants repeat steps 4-7 with a second swab — once completed, add the participant's swab to the 50 ml Pooled Sample tube. All participants should have one swab in the same 50 ml tube. Tightly seal cap.
9. Ensure each participant has placed one (1x) swab into the shared 50mL Pooled Sample tube and one (1x) swab into their own 10mL individual tube.

(2x) Samples Per Participant





Perimeter

03

Register tests

1. Log in to the Perimeter Platform at

testcenter.concentricbyginkgo.com/login.

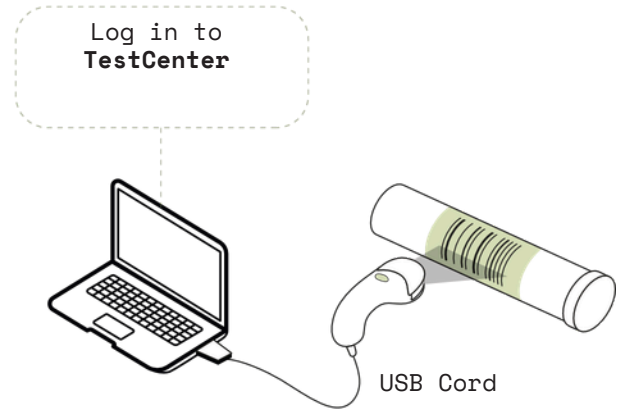
2. Connect the barcode scanner to your computer (for the best user experience, use the provided USB cord to connect the barcode scanner to a laptop instead of using Bluetooth).

3. Log the pooled test in TestCenter by scanning the barcode.

- Name the pool and indicate how many participants (must be between 5 and 10) are in each pool.

4. Add each participant to the pooled test and log their individual test in the Concentric Testing platform.

- In the **“Add a Test Taker”** field, type the participant’s confirmation number, confirming the date and country of origin.
- Click **“Add to Pool”** to add the Test Taker and individual sample to the pool.
- Scan the individual barcode under **“Double Collection Tube ID,”** making sure to match the correct confirmation number and individual barcode.



04

Prepare for shipment

1. Place each participant’s 10mL sample tube inside one of the zip-lock “biohazard” bags.

2. Make sure that individual tubes are closed and not leaking before adding to the biohazard bags. If the caps are too tight, or not well positioned, the liquid will leak, contaminating the other samples and invalidating the test.

3. Place the shared 50mL “pool” sample tube inside the other zip-lock biohazard bag.

- 50mL pool tubes should never be in the same biohazard bag as 10mL individual tubes.

4. Ensure the number of swabs in the pool matches the number of individual tests registered.

5. Ensure both biohazard bags are labeled with the appropriate matching “pool” barcode label and are properly sealed + labeled.

6. Place a single rubber band around both the “pool” biohazard bag and the “individual” biohazard bag so they are received by the lab in unison.

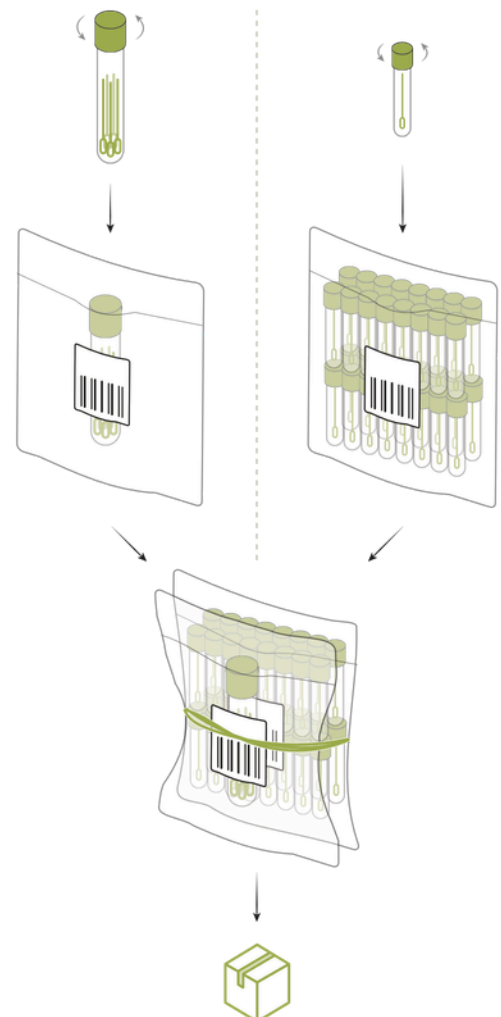
7. Once completed, place the rubber-banded biohazard bags inside of the intended shipping box.

8. Safely pack as many pools into a single shipping box as reasonably fit. Overloading a single box may jeopardize sample integrity.

9. Ship all samples back to the laboratory in an appropriately labeled UN3373 shipping box (boxes provided through the program will already be labeled).

10. Follow the shipping instructions provided by Perimeter.

11. Ship older samples first if the courier cannot accept all samples.



a. *Note: samples are stable for up to 72 hours (3 days) at room temperature followed by up to 48 hours (2 days) refrigerated (2-8°C)*